



The Effect of Inclusion Level of Soybean Oil and Palm Oil on Their Digestible and Metabolizable Energy Content Determined with the Difference and Regression Method When Fed to Growing Pigs

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ABSTRACT: This experiment was conducted to determine the effects of inclusion level of soybean oil (SO) and palm oil (PO) on their digestible and metabolism energy (DE and ME) contents when fed to growing pigs by difference and regression method. Sixty-six crossbred growing barrows (Duroc×Landrace×Yorkshire and weighing 38.1±2.4 kg) were randomly allotted to a 2×5 factorial arrangement involving 2 lipid sources (SO and PO), and 5 levels of lipid (2%, 4%, 6%, 8%, and 10%) as well as a basal diet composed of corn and soybean meal. The barrows were housed in individual metabolism crates to facilitate separate collection of feces and urine, and were fed the assigned test diets at 4% of initial body weight per day. A 5-d total collection of feces and urine followed a 7-d diet adaptation period. The results showed that the DE and ME contents of SO and PO determined by the difference method were not affected by inclusion level. The DE and ME determined by the regression method for SO were greater compared with the corresponding respective values for PO (DE: 37.07, ME: 36.79 MJ/kg for SO; DE: 34.11, ME: 33.84 MJ/kg for PO, respectively). These values were close to the DE and ME values determined by the difference method at the 10% inclusion level (DE: 37.31, ME: 36.83 MJ/kg for SO; DE: 34.62, ME: 33.47 MJ/kg for PO, respectively). A similar response for the apparent total tract digestibility of acid-hydrolyzed ether extract (AEE) in lipids was observed. The true total tract digestibility of AEE in SO was significantly ($p<0.05$) greater than that for PO (97.5% and 91.1%, respectively). In conclusion, the DE and ME contents of lipid was not affected by its inclusion level. The difference method can substitute the regression method to determine the DE and ME contents in lipids when the inclusion level is 10%. (**Key Words:** Digestibility, Energy Content, Inclusion Level, Palm Oil, Pigs, Soybean Oil)

INTRODUCTION

Lipids are an important dietary energy source with a high energy content and high digestibility for swine (Li et al., 1990; Jone et al., 1992; Jørgensen and Fernandez, 2000). An accurate determination of the energy content of a lipid is important for utilizing it appropriately in the feed industry. The determined energy values for the same source of lipid in different studies are not consistent (Wiseman et al., 1990; Powles et al., 1993; Jørgensen et al., 2000). In addition to breed of animal, basal diet, and determination method, the

inclusion level of the lipid is an important factor that affects the accuracy of the determination of the energy value of a lipid (Allee et al., 1971; Kil et al., 2011).

The extensively used prediction equation for the energy value of lipids is based on free fatty acids content and degree of saturation. However, increasing the inclusion level of lipid can affect the values for the digestibility of acid-hydrolyzed ether extract (AEE), dry matter (DM), crude protein (CP), and some amino acids in growing pigs (Jørgensen et al., 2000; Albin et al., 2001; Bruce et al., 2006; Rojas-Cano et al., 2014). Therefore, there is a hypothesis that interactions exist between the basal diet and the supplemented lipid which result in changes in the determined energy value of the lipid. To our knowledge,

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there is limited information about systematic evaluation of the energy value of different dietary lipids for pigs.

The composition of the fatty acids in lipids may affect their digestibility, such as saturated or unsaturated lipid (Wiseman et al., 1990; Powles et al., 1993; Jørgensen and Fernandez, 2000; Kil et al., 2011). In addition, the effect of inclusion level on the digestibility and energy value of a saturated or unsaturated lipid might be different. Therefore, the objective of this study was to determine the effect of inclusion level of soybean oil (SO) and palm oil (PO) on their digestible energy (DE) and metabolism energy (ME) contents determined by difference and regression method.

MATERIALS AND METHODS

General procedures

The experimental protocol used in this study was approved by the Institutional Animal Care and Use Committee at China Agricultural University (Beijing, China). This study was conducted in the Metabolism Laboratory of the National Feed Engineering Technology Research Center (HeBei Province, China).

The barrows were housed individually in stainless steel metabolism cages (1.4×0.7×0.6 m) equipped with a feeder and a nipple drinker. The crates were located in six environmentally controlled rooms with the temperature maintained at 22°C to 24°C. To minimize the opportunity for environmental effects to confound the results, one pig per treatment was allotted to each room. The pigs were allowed a 10 day period to adapt to the metabolism crates

and the environmental conditions of the room. During this period, the pigs were fed a commercial diet and feed intake was gradually increased until the daily feed allowance reached 4% of the pig's body weight (Adeola, 2001). Pigs were weighed at the beginning of the adaptation period.

Sixty-six crossbred growing barrows (Duroc×Landrace ×Yorkshire and weighing 38.1±2.4 kg) were randomly allotted to a 2×5 factorial arrangement involving 2 lipid sources (SO and PO), and 5 levels of lipid (2%, 4%, 6%, 8%, and 10%) as well as a basal diet composed of corn and soybean meal (Table 1). Basal diet was substituted for lipid at graded levels (2%, 4%, 6%, 8%, and 10%) according to the difference procedure (Kong and Adeola, 2014). The SO and PO were food grade (FFA<0.2%). The fatty acid profile of the lipids was presented in Table 2.

All experimental diets were supplemented with the same proportion of dicalcium phosphate, limestone, salt, and vitamin-mineral premix to meet or exceed the estimated nutrient requirements for growing pigs as recommended by NRC (2012).

Feed was provided in mash form twice daily at 08:00 and 17:00 h. Water was available continuously through a low-pressure nipple drinker. A 5-d period for total collection of feces and urine followed a 7-d period for diet adaptation. Feed remnants and feed spillage were collected, dried and weighed accurately to calculate feed intake.

Feces were collected immediately as they appeared in the metabolism crates and placed in plastic bags to be stored at -20°C. Urine was collected in a bucket placed under the metabolism crate. The bucket contained 10 mL of 6 N HCl

Table 1. Ingredient composition and chemical analysis of the experimental diets (% as-fed)

Level of added oil (%)	Soybean oil						Palm oil				
	0	2	4	6	8	10	2	4	6	8	10
Ingredient (%)											
Corn	75.40	73.85	72.30	70.76	69.21	67.66	73.85	72.30	70.76	69.21	67.66
Soybean meal	22.00	21.55	21.10	20.64	20.19	19.74	21.55	21.10	20.64	20.19	19.74
Soybean oil	-	2.00	4.00	6.00	8.00	10.00	-	-	-	-	-
Palm oil	-	-	-	-	-	-	2.00	4.00	6.00	8.00	10.00
Dicalcium phosphate	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90
Limestone	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90	0.90
Salt	0.30	0.30	0.30	0.30	0.30	0.30	0.30	0.30	0.30	0.30	0.30
Vitamin-mineral premix ¹	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
Analyzed composition (%)											
Dry matter	87.83	88.12	88.34	88.54	88.84	89.15	88.17	88.34	88.55	88.81	89.05
Acid-hydrolyzed ether extract	2.91	4.98	7.09	9.01	10.55	12.57	5.32	7.21	9.13	10.84	12.78
Crude protein	16.06	15.86	15.35	15.11	14.96	14.62	15.83	15.55	15.11	15.06	14.41
Neutral detergent fiber	8.35	8.12	7.72	7.79	7.07	7.32	8.16	8.18	7.55	7.64	7.06
Acid detergent fiber	2.43	2.38	2.31	2.29	2.14	2.13	2.60	2.48	2.31	2.20	2.31
Ash	4.36	4.25	4.32	4.12	4.26	4.16	4.50	4.26	4.21	4.27	4.11

¹ Premix provided the following per kg of complete diet for growing pigs: vitamin A, 5,512 IU; vitamin D₃, 2,200 IU; vitamin E, 30 IU; vitamin K₃, 2.2 mg; vitamin B₁₂, 27.6 µg; riboflavin, 4 mg; pantothenic acid, 14 mg; niacin, 30 mg; choline chloride, 400 mg; folacin, 0.7 mg; thiamine 1.5 mg; pyridoxine 3 mg; biotin, 44 µg; Mn, 40 mg (MnO); Fe, 75 mg (FeSO₄·H₂O); Zn, 75 mg (ZnO); Cu, 100 mg (CuSO₄·5H₂O); I, 0.3 mg (KI); Se, 0.3 mg (Na₂SeO₃).

Table 2. The fatty acid profile of lipids

Item	Soybean oil	Palm oil
Fatty acid (% of lipid)		
C8:0	0.13	0.13
C12:0	0.01	0.13
C14:0	0.10	0.88
C16:0	11.49	46.16
C16:1	0.09	0.13
C18:0	4.33	5.07
C18:1	23.67	37.18
C18:2	52.80	9.08
C18:3	5.77	0.36
C20:0	0.41	0.41
C20:1	0.21	0.10
C22:0	0.45	0.08
C24:0	0.17	0.08
Others	0.35	0.23
SFA	17.34	53.12
MUFA	24.03	37.43
PUFA	58.62	9.44

SFA, saturated fatty acid; MUFA, monounsaturated fatty acid; PUFA, polyunsaturated fatty acid.

for every 1,000 mL of urine. Each day, the total urine volume was measured and a 10% aliquot was filtered through gauze and the urine samples were transferred into a screw-capped tube and immediately stored at -20°C until needed for analysis. At the end of the collection period, feces were thawed, pooled, homogenized, sub-sampled, dried for 72 h at 65°C in a forced-air drying oven and ground through a 1-mm screen before chemical analysis.

Chemical analysis

All chemical analyses were conducted in duplicate and repeated if the results differed by more than 5%. Feed and fecal samples were analyzed for DM (AOAC method 934.01), AEE (AOAC method 954.02), CP (AOAC method 990.03), crude fiber (AOAC method 978.10), and ash (AOAC method 942.05). The content of neutral detergent fiber (NDF) and acid detergent fiber (ADF) were determined using filter bags and fiber analyzer equipment (Fiber Analyzer, Ankom Technology, Macedon, NY, USA) following a modification of the procedure of Van Soest et al. (1991). The content of gross energy (GE) was determined using an Automatic Adiabatic Oxygen Bomb Calorimeter (Parr 6300 Calorimeter, Moline, IL, USA). The fatty acid profiles of the lipid sources were determined by gas chromatography (6890 series, Agilent Technologies, Wilmington, DE, USA) following the procedures of Sukhija and Palmquist (1988).

Calculations and statistical analyses

The apparent total tract digestibility (ATTD) of AEE,

DM, NDF, ADF, and CP were calculated for each diet. The true total tract digestibility (TTTD) of lipid was calculated using the regression method. The apparent digested fat (g/kg of dry matter intake [DMI]) of the entire intestinal tract was regressed against dietary fat intake (g/kg DM) for each pig. The slope of the regression line represented the true digestibility of lipid, whereas the Y-intercept of this regression equation was considered the endogenous loss of AEE (g/kg of DMI) (Jørgensen et al., 1993).

The DE and ME values of SO were calculated using the following equations:

$$\text{DE}_{\text{lipid}} = \frac{\text{wt. feed intake} \times \text{DE}_{\text{feed}} - \text{wt. EYI intake} \times \text{DE}_{\text{EYI}}}{\text{wt. lipid intake}}$$

$$\text{ME}_{\text{lipid}} = \frac{\text{wt. feed intake} \times \text{ME}_{\text{feed}} - \text{wt. EYI intake} \times \text{ME}_{\text{EYI}}}{\text{wt. lipid intake}}$$

In these equations, wt. = weight and EYI = energy-yielding ingredient of basal diet. Because the proportion of the EYI in basal diets was 0.974, the energy concentration of EYI was calculated by dividing the DE or ME of the basal diet by 0.974.

The ATTD of lipid contained in each experimental diet was calculated using the following equation:

$$\text{D}_{\text{lipid}} = \frac{(\text{wt. feed intake} \times \text{C}_{\text{AEE feed}} \times \text{D}_{\text{AEE feed}}) - (\text{wt. EYI intake} \times \text{C}_{\text{AEE EYI}} \times \text{D}_{\text{AEE EYI}})}{\text{wt. lipid intake}}$$

In this equation, C_{AEE} = the concentration of AEE and D_{AEE} = the digestibility of AEE.

The UNIVARIATE procedure of SAS was used to check for normal distribution of the model residuals and equal variances. The residual vs the predicted plot procedure was used to identify outliers. The model included the effect of basal diet, inclusion level of SO and their interaction for all measurements. Data were analyzed using the MIXED procedure of SAS (SAS Inst. Inc., Cary NC, USA). The individual animal and dietary treatment were the experimental units for analyzing the data from the digestibility trial. The model included diet as a fixed effect and animal as a random effect. Orthogonal polynomial contrasts were used to detect linear and quadratic responses to inclusion level of lipid. The REG procedure of SAS was used to estimate the Y-intercept of the regression line for determining endogenous loss of AEE, and the slope was

Table 3. The digestible (DE) and metabolizable energy (ME) contents of the experimental diets (as-fed basis)¹

Item	Inclusion level (%)						SEM	p-value		
	0	2	4	6	8	10		Level	Source	Interaction
DE (MJ/kg)										
Soybean oil ²	14.15	14.47	14.94	15.39	15.92	16.42	0.142	<0.001	0.022	0.324
Palm oil ²	14.15	14.46	14.91	15.33	15.75	16.15	0.127			
ME (MJ/kg)										
Soybean oil ²	13.87	14.19	14.64	15.08	15.56	16.13	0.148	<0.001	0.075	0.370
Palm oil ²	13.87	14.18	14.62	15.02	15.43	15.80	0.116			
ME/DE (%)										
Soybean oil	98.08	98.06	98.09	97.98	97.93	98.23	0.120	0.608	0.995	0.924
Palm oil	98.08	98.05	98.02	97.98	97.94	97.79	0.109			

SEM, standard error of the mean.

¹ n = 6 per mean. ² Linear effect of inclusion level at p<0.001.

used to determine the TTTD for lipid. Intercepts and slopes were compared between the two basal diets based on confidence intervals coefficients for regression lines (Dilger and Adeola, 2006). The level of significance adopted was 5% (p<0.05) to assess differences among means.

RESULTS

All pigs remained healthy and readily consumed their diets without any problems. Both feces and urine were successfully collected from all pigs. The DE and ME contents of the experimental diets linearly increased (p<0.001) as inclusion level increased regardless of lipid source (Table 3). The lipid source significantly (p = 0.022) affected the DE content of the experimental diets but did not affect the ME content of the experimental diets (p = 0.075). The average ratio of ME to DE for diets containing lipid was 98%.

At least two outliers were presented in the 2% inclusion level treatment for DE and ME of lipids. The coefficient of variance of DE and ME contents within replications decreased inversely as the inclusion level of lipid increased

(p<0.01, R²>0.9, Figure 1). The coefficients of variance for the DE and ME contents of lipids at the 2% inclusion level were greater than 23% after excluding the outliers. This was due to the calculation method of difference method. The stand error of energy value for 2% inclusion level (not shown in Table 4) was also much greater than other higher level treatments, which was similar to the result of Villamied (1996). Therefore, in the process of data analysis, it was inappropriate to analyze the effect of inclusion level on DE or ME of lipids determined by difference method including 2% level treatment.

The determined DE and ME contents of SO and PO by the difference method were not affected by inclusion level (Table 4). The lipid source significantly affected the DE content of lipids (p<0.05). However, the ME content was not affected by lipid source (p = 0.145). There was no effect of inclusion level and lipid source on the ratio of ME/DE for lipids. The ME was 97.73% and 97.07% of DE for SO and PO respectively, which are close to the recommended value 98% in NRC (2012).

The DE and ME contents of SO and PO were also determined by the regression method. The values of DE and

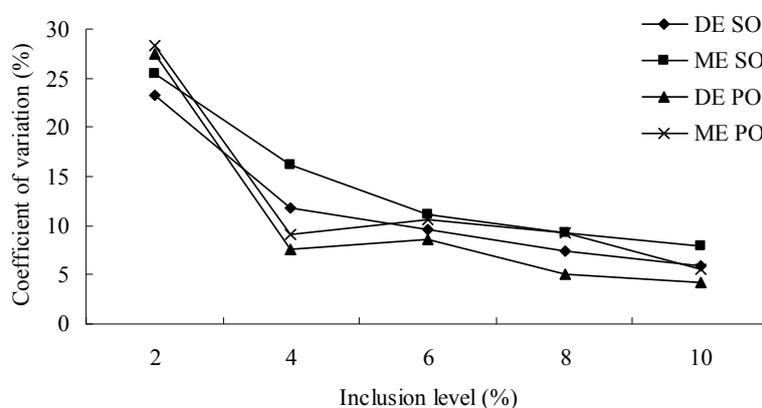


Figure 1. Effect of the inclusion level on the coefficient of variation of the determined DE and ME contents of lipids. n = 5. The coefficient of variance of DE and ME contents within replications decreased inversely as the inclusion level of lipids increased (p<0.01, R²>0.9). DE, digestible energy, ME, metabolism energy, SO, soybean oil; PO, palm oil.

Table 4. The digestible (DE) and metabolizable energy (ME) contents of soybean oil and palm oil determined by the difference method in growing pigs^{1,2}

Item	Inclusion level (%)				Mean value	SEM	p-value		
	4	6	8	10			Level	Source	Interaction
DE (MJ/kg)									
Soybean oil	34.51	35.25	36.74	37.31	35.95	0.650	0.345	0.047	0.816
Palm oil	33.63	34.31	34.59	34.62	34.27	0.447			
ME (MJ/kg)									
Soybean oil	33.35	34.29	35.26	36.83	34.99	0.857	0.573	0.145	0.787
Palm oil	32.81	33.38	33.65	33.47	33.34	0.582			
ME/DE (%)									
Soybean oil	97.66	97.24	97.32	98.63	97.73	1.031	0.994	0.649	0.953
Palm oil	97.33	97.18	97.13	96.70	97.07	0.740			

SEM, standard error of the mean.

¹ n = 6 per mean.² The 2% inclusion level was not used to analyze the effect of inclusion level on the DE and ME contents of lipids, because at least two outliers were presented in the 2% inclusion level treatment for DE and ME of lipids, and the coefficients of variation were still greater than 23% after excluding the outliers.

ME for SO were greater compared with the corresponding values for PO (DE: 37.07, ME: 36.79 MJ/kg for SO; DE: 34.11, ME: 33.84 MJ/kg for PO, respectively, Table 5). The values of DE and ME for SO and PO derived from the regression method were not significantly different from the values derived from the difference method determined by a *t*-test, so that the result was not shown in a Table 5.

The ATTD of AEE in SO and PO for all inclusion levels were calculated in the present experiment. Likewise, it was similar to the DE and ME of lipids, the 2% level treatment was also not used to analyze the effect of inclusion level on the ATTD of AEE in lipid determined by difference method.

As the inclusion level increased, the ATTD of AEE in SO increased, while the ATTD of AEE in PO decreased, but the differences were not statistically significant (Table 6). The inclusion level of the dietary lipids did not affect the ATTD of AEE in lipids, but there was a significant difference ($p = 0.002$) between the ATTD of AEE in SO and PO.

The regression of total tract digested AEE against the intake of AEE was significant for SO and PO ($p < 0.001$, $R^2 > 0.99$, Table 7). The TTTD of AEE in SO, calculated from the slope of the regression equation, was significantly greater ($p < 0.001$) than that for PO (97.5% and 91.1%, respectively). The *y*-intercepts that represented the

Table 5. The digestible (DE) and metabolizable energy (ME) of soybean oil and palm oil determined by the regression method¹

Item	Regression equation ²	R ²	Intercept		Slope		When x = 1,000, DE/ME
			SE	p-value	SE	p-value	
DE (MJ/kg)							
Soybean oil	$y = 0.023x + 14.07$	0.946	0.060	<0.001	0.001	<0.001	37.07
Palm oil	$y = 0.020x + 14.11$	0.961	0.045	<0.001	0.001	<0.001	34.11
ME (MJ/kg)							
Soybean oil	$y = 0.023x + 13.79$	0.920	0.076	<0.001	0.001	<0.001	36.79
Palm oil	$y = 0.020x + 13.84$	0.932	0.060	<0.001	0.001	<0.001	33.84

SE, standard error.

¹ n = 36.² These linear regression equations are all significant ($p < 0.001$). In equation $y = ax + b$, $y =$ DE or ME values of diet (MJ/kg), $x =$ inclusion level of lipid (g/kg), and $b =$ intercept.**Table 6.** The apparent total tract digestibility (ATTD) of acid-hydrolyzed ether extract (AEE) in soybean oil and palm oil (%)^{1,2}

Item	Inclusion level (%)				Mean value	SEM	p-value		
	4	6	8	10			Level	Source	Interaction
Soybean oil	93.12	93.87	94.47	95.83	94.32	0.649	0.997	0.002	0.260
Palm oil	92.57	91.43	91.06	90.03	91.27	0.636			

SEM, standard error of the mean.

¹ n = 6 per mean.² The 2% inclusion level was not used to analyze the effect of inclusion level on ATTD of lipids, because at least two outliers were presented in the 2% inclusion level treatment of lipids, and the stand error was much greater than the other levels.

Table 7. The true total tract digestibility (TTTD) and the endogenous loss (EL) of acid-hydrolyzed ether extract (AEE) for soybean oil and palm oil determined through the regression of total apparently digested AEE (g/kg of DMI) against the dietary AEE intake (g/kg of DM)¹

Item	Regression equation	R ²	EL of AEE (g/kg of DMI)			TTTD (%)			ATTD (%) ⁴
			Estimate	SE	p-value ²	Estimate	SE	p-value ³	
Soybean oil	y = 0.975x-14.02	0.996	14.02	1.121	<0.001	97.5 ^a	0.011	<0.001	96.1 ^a
Palm oil	y = 0.911x-10.80	0.997	10.80	0.916	<0.001	91.1 ^b	0.009	<0.001	90.0 ^b

DMI, dry matter intake; DM, dry matter, SE, standard error; ATTD, apparent total tract digestibility.

¹ n = 36.

² The probability of significance for the intercept of the regression equation.

³ The probability of significance for the slope of the regression equation.

⁴ The apparent total tract digestibility, calculated as the x = 1,000, ATTD = y/1,000.

^{a-b} Within a column means followed by different letters are different (p<0.05).

endogenous losses for AEE of SO and PO were 14.02 and 10.80 g/kg of DMI respectively (p<0.001). However, there was no difference between them. The ATTD of AEE in lipids were also calculated from these equations. The ATTD AEE in was greater for SO than PO (96.1% and 90.0%, respectively, p<0.05).

A linear increase (p<0.05, Table 8) in the ATTD of GE, DM, CP, and AEE and a quadratic increase (p<0.01) in the ATTD of AEE were observed in pigs fed diets containing SO as inclusion level increased. However, there was no effect of inclusion level on the ATTD of NDF and ADF in the diet containing SO. A quadratic increase (p<0.05) in the ATTD of DM and a linear increase (p<0.01) in the ATTD of AEE were observed in pigs fed diets containing PO as the

inclusion level increased. However, no differences in the ATTD of GE, CP, NDF, and ADF were observed due to inclusion level. Lipid source affected the ATTD of GE and AEE for pigs fed diets containing SO or PO (p<0.01).

DISCUSSION

A linear increase in the ATTD of AEE for the experimental diets containing SO or PO with increasing inclusion level was observed and is in agreement with previous results from research with SO (Jørgensen et al., 1993; Kil et al., 2011; Adeola et al., 2013), corn oil (Kil et al., 2010), tallow (Adeola et al., 2013), poultry fat (Mendoza and Heugten, 2014) and rapeseed oil (Jørgensen

Table 8. The apparent total tract digestibility of dry matter (DM), crude protein (CP), acid-hydrolyzed ether extract (AEE), neutral detergent fiber (NDF), and acid detergent fiber (ADF) of each experimental diet (%)¹

Item	Inclusion level (%)						SEM	p-value		
	0	2	4	6	8	10		Level	Source	Interaction
DM										
Soybean oil ²	88.75	88.22	88.72	89.12	89.66	90.34	0.212	0.001	0.082	0.829
Palm oil ³	88.75	88.12	88.46	88.86	88.95	89.44	0.154			
GE										
Soybean oil ²	88.35	88.01	88.56	89.13	89.84	90.45	0.235	0.002	0.003	0.308
Palm oil	88.35	87.92	88.13	88.51	88.46	88.81	0.154			
CP										
Soybean oil	87.80	87.44	87.62	87.96	88.64	89.17	0.244	0.169	0.280	0.873
Palm oil	87.80	87.34	87.57	87.81	88.03	88.14	0.178			
AEE										
Soybean oil ^{2,3}	59.45	72.54	78.54	82.44	86.02	88.38	1.710	<0.001	<0.001	0.067
Palm oil ²	59.45	72.29	77.81	80.46	82.77	83.38	1.458			
NDF										
Soybean oil	52.25	46.53	52.34	52.41	53.08	52.71	0.906	0.141	0.113	0.983
Palm oil	52.25	49.75	53.20	55.89	55.30	55.67	0.985			
ADF										
Soybean oil	49.27	44.51	52.91	49.83	50.88	50.59	1.178	0.607	0.062	0.592
Palm oil	49.27	52.42	51.27	53.61	53.54	57.23	1.268			

SEM, standard error of the mean; GE, gross energy; CP, crude protein.

¹ n = 6 per mean.

² Linear effect of inclusion level at p<0.05.

³ Quadratic effect of inclusion level at p<0.05.

et al., 1996). Likewise, DE, ME and ATTD of DM and GE in diets increased with increasing inclusion level of lipids. This is due to the greater GE, DE, and ME contents and higher digestibility of AEE in lipid sources than in corn and soybean meal.

Observations of greater DE content and relatively greater ME content in diets containing SO than in diets containing PO, found in the current study, are not consistent with previous research which compared diets containing SO or tallow fed from d 21 to 35 postweaning (Adeola et al., 2013). The basis for this discrepancy is unclear but may be that the tallow had higher digestibility because of the position and distribution of saturated and unsaturated fatty acids on the triglyceride molecules within the tallow (Bracco, 1994; Ramírez et al., 2001).

In the current study, the DE and ME contents of lipids were determined at every inclusion level of lipids by the difference method. The DE and ME contents of the lipid sources were not affected by the dietary lipid level. However, the DE content of lipids was significantly affected by lipid source. The reason for the observation may be the greater digestibility of SO than PO. The ratio of unsaturated fatty acid to saturated fatty acid can affect the DE and ME contents of lipid (Wiseman et al., 1990; Powles et al., 1993; Jørgensen et al., 2000). However, lipid source did not affect the ME content of lipids in the current study, which was an interesting finding. Perhaps lipid source can not affect the retention of protein for pigs (Kil et al., 2011).

Increasing the dietary lipid level increased the DE and ME contents of lipids, even though the difference was not significant. The DE and ME contents of SO increased to a greater extent with increasing inclusion level compared with PO. The observation in the DE and ME response is consistent with the observation for the ME content of diets containing SO or tallow (Adeola et al., 2013). That may be due to the changing of the ratio of unsaturated to saturated fatty acids of the dietary lipid in diets with increasing inclusion level. Along with the increased inclusion level of lipid, dietary lipids containing more unsaturated fatty acids can yield more available energy compared with dietary lipids containing more saturated fatty acids (Stahly, 1984; Powles et al., 1995).

The DE and ME of lipids determined with the regression equations were not statistically different from the values determined with the difference method, and were very close to the values at the highest level regardless of lipid sources (unsaturated and saturated). The regression method is largely used to determine the energy content of lipids (Wiseman et al., 1990; Powles et al., 1993; Baidoo et al., 1996; Jørgensen and Fernandez, 2000). However, it is time-consuming and very costly. From the results, we found the coefficient of variance of the determined DE and ME values were small when the inclusion level was 10%.

Therefore, the difference method with a 10% inclusion level can be used to determine the DE and ME contents of lipids for growing pigs.

General theory considers that a greater lipid digestibility is usually in response to a greater DE content for lipid. Previous studies evaluated the DE content of lipids through multiplying the GE by the ATTD of lipid (Wiseman et al., 1990; Powles et al., 1993). However, the contrary results of the current study obtained from difference method showed that the digestibility of the supplemental lipid might be affected by intact fat or other nutrients in the basal diet. Soybean oil is rich in unsaturated fatty acids. However, PO is rich in saturated fatty acids. As the inclusion level of SO increased, the concentration of unsaturated fatty acids in the experimental diets will increase. Likewise for PO, the concentration of saturated fatty acids in the experimental diets will increase. Because unsaturated fatty acids have a greater potential for micelle formation than saturated fatty acids, unsaturated fatty acids are more digestible than saturated fatty acids (Freeman et al., 1968; Stahly, 1984; Powles et al., 1995).

The concentration of apparently digested AEE in the basal diet was about 17 g/kg in the current study. The digested AEE mainly was corn oil containing more unsaturated fatty acids (82.3%, NRC, 2012). As the inclusion level of PO increased, the ratio of unsaturated fatty acids to saturated fatty acids in the experimental diets will decrease. This is probably the reason for the ATTD of AEE in PO being lower at a higher lipid level in the current study. Unsaturated fatty acids may aid in the digestion of saturated fatty acids by increasing micelle formation of saturated fatty acids, and the digestibility of saturated fatty acid is improved if unsaturated fatty acids are mixed with saturated fatty acids (Powles et al., 1993).

The values for endogenous loss of AEE for SO and PO were greater than previous values reported for SO (Jørgensen et al., 1993) or corn oil (Kil et al., 2010; Kim et al., 2013). The value for SO was slightly greater than that for PO, but there was no significant difference between them. This might be because SO with a high content of unsaturated fatty acids can facilitate the excretion of endogenous loss in the feces due to the microbial hydrogenation of unsaturated fatty acids in the hindgut (Jørgensen et al., 1993).

The greater ATTD and TTTD of AEE derived from the regression equation in SO compared with PO that was observed in the current study is in agreement with previous studies (Powles et al., 1993; Jørgensen et al., 2000). This observation is attributed to the greater digestibility of unsaturated lipid compared with saturated lipid. It was interesting to find that the ATTD of AEE in lipids derived from the regression equation was also very close to the value at the highest level in the present experiment. The

observation for the AEE in lipids digestibility response is consistent with the observation for the DE and ME response.

CONCLUSION

The DE content, TTTD, and ATTD of AEE in lipids can be affected by lipid sources. The DE and ME contents in lipids were not affected by inclusion level in the range of 0% to 10%. Therefore, the difference method can substitute the regression method to determine the DE and ME contents of lipids when the inclusion level is 10%.

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