



Effects of Supplemental Mannanligosaccharides on Growth Performance, Faecal Characteristics and Health in Dairy Calves

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ABSTRACT: Twenty Holstein calves were used to investigate the effects of mannanligosaccharides (MOS) supplementation in the whole milk on growth performance, faecal score, faecal pH, selected faecal bacterial populations and health during the preweaning period. Healthy calves selected by clinical examination were allocated to one of the two groups (control [CG] and experimental [EG]) at 5 days old. Each group consisted of 5 male and 5 female calves. Each calf in EG was supplemented with 7 g/d of a MOS product (Celmanax) from 5 days to 56 days of age. MOS supplement was mixed with the whole milk once in the morning and administered to the calves in EG via nipple bottle, whereas the calves in CG were fed the whole milk without MOS. Calves were weaned at 56 days of age. The final body weight, average daily weight gain (ADG) and average daily feed intake (ADFI) were statistically similar ($p > 0.05$) but were higher by 3.70%, 6.66%, and 10.97%, respectively, in MOS than in control calves. Feed efficiency (ADG/ADFI) was also similar in two calves group. While faecal scores did not differ on day 5, 7, 14, 21, 28, 42, 49, and 56 between groups, EG had a higher faecal score ($p = 0.05$) than CG on day 35. Faecal concentration of *Lactobacillus* was lower ($p < 0.05$) in EG compared with CG. No differences ($p > 0.05$) in faecal concentrations of *Bifidobacterium*, *Clostridium perfringens*, and *Escherichia coli* were found between groups. Although there were no significant differences ($p > 0.05$) in the incidence of diarrhoea, treatment days for diarrhoea and the costs associated with diarrhoea treatments between groups, collectively, the observed reductions in treatment days and the cost of diarrhoea treatments accompanying increases in final body weight, ADG and ADFI for EG may indicate potential benefit of MOS in treatment of diarrhoea. (**Key Words:** Calves, Mannanligosaccharides, Growth Performance, Faecal Characteristics, Health)

INTRODUCTION

The maintenance of calf health and optimizing calf growth are key objectives especially at early stages of life (Ghosh and Mehla, 2012). Intestinal infections and subsequent diarrhoea account for the majority of health problems affecting calves during the preweaning period and are the primary reason for death and poor growth performance in the first 2 months of age (Davis and Drackley, 1998). In this respect, prebiotics have been used to improve the health and performance of calves during the preweaning period (Heinrichs et al., 2003; Ghosh and Mehla, 2012; Heinrichs et al., 2013).

Prebiotics are defined as nondigestible carbohydrates that can be able to beneficially modify the growth and/or activity of the intestinal bacterial populations towards a healthier microflora and improve indices of host health (Gibson and Roberfroid, 1995). Fructooligosaccharides (Grand et al., 2013), galactooligosaccharides (Kanakupt et al., 2011) and mannanligosaccharides (MOS) (Demirel et al., 2007; Ghosh and Mehla, 2012) have been utilized as prebiotic in animals. Supplementation of MOS has become relatively common throughout the world. This substance is available for inclusion in milk, milk replacer and calf starter (Ghosh and Mehla, 2012). MOS are complex mannose sugars derived from the cell wall of *Saccharomyces cerevisiae* (Spring et al., 2000). The structure of the mannan component of MOS resembles that of the carbohydrates on the intestinal wall. Pathogenic bacteria containing type-I (mannose specific) fimbriae normally adhere to mannans on

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the mucosal surface of the intestine. The mannan component of MOS provides a competitive binding site for certain intestinal pathogens (Newman, 1994; Spring et al., 2000). Therefore, benefits of MOS are associated with pathogen removal from the intestine without attachment and colonization (Spring et al., 2000; Shane, 2001).

In some studies with calves, supplementation with MOS has been shown to increase average daily weight gain (ADG) and feed efficiency (ADG/ADFI [average daily feed intake]) (Ghosh and Mehla, 2012; Heinrichs et al., 2013), to improve faecal score (Heinrichs et al., 2003; Morrison et al., 2010), and to decrease faecal concentrations of bacterial populations such as total coliform and *Escherichia coli* (Jacques and Newman, 1994; Ghosh and Mehla, 2012). However, other data indicated that supplementation of MOS did not lead to the improvement in growth performance (Hill et al., 2008; Hill et al., 2009) and faecal score (Terre et al., 2007; Uzmay et al., 2011) and the change in faecal bacterial populations (Terre et al., 2007) in calves. Heinrichs et al. (2003) reported that addition of MOS to milk replacer reduced the incidence and severity of diarrhoea in calves. Conversely, Hill et al. (2008) and Uzmay et al. (2011) observed that feeding milk replacer or whole milk containing MOS did not reduce diarrhoea and days with abnormal faecal scores (diarrhoea) in calves.

Results from feeding MOS are variable and are not yet fully conclusive. Because of these different results, further research efforts should continue to evaluate the efficacy of MOS on performance and health in calves. The objective of the current study was to investigate the effects of MOS supplementation on growth performance, faecal score, faecal pH, selected faecal bacterial populations (*Lactobacillus*, *Bifidobacterium*, *Clostridium perfringens*, and *E. coli*) and health in calves.

MATERIALS AND METHODS

Study area and animals

The study was carried out at Uludag University Applied Research Center for Veterinary Faculty Unit in Bursa located within the North West Turkey. This study was conducted under an approved protocol by Animal Care and Use Committee of University of Uludag.

Twenty healthy calves (5 days old) of ten each male and female were selected by clinical examination (body temperature, respiration, and pulsation rates), general appearance and occurrence of diarrhoea. Calves were randomly divided into two equal groups (control [CG] and experimental [EG]) of five male and five female.

Management and experimental design

Calves were fed colostrum within 2 h of birth and

housed in individual pens until the end of the study at 56 days of age. Calves received 4 L/d of the whole milk from day 2 to 49 and 2 L/d of the whole milk during last 7 days of the study. Milk was offered twice daily. Pelleted calf starter and water were given *ad libitum* from day 5 (the beginning of the study). Pelleted calf starter was containing 19.58% of crude protein, 5.7% of crude fat, 22.46% of neutral detergent fiber, 9.99% of acid detergent fiber, 3.55% of acid detergent lignin, 43.23% of non-fiber carbohydrate and 9.03% of ash on a dry matter basis. Calves were weaned at 56 days of age.

Each calf in EG was supplemented with 7 g/d of a MOS product from 5 days to 56 days of age. MOS supplement was mixed with the whole milk once in the morning and administered to the calves in EG via nipple bottle at the amount recommended by the manufacturer (Celmanax, Vi-COR, Mason City, IA, USA). The prebiotic product obtained from cell wall fragments of *S. cerevisiae* contained 51% of glucan and 25.5% of mannan according to the certificate of analysis reported by Vi-COR.

Measurements and sample collection

Individual pelleted calf starter intake was measured daily by collecting residual feed after every feeding during the study. Calves were weighed before feeding at day 5 and 56. ADG, ADFI, and feed efficiency (ADG/ADFI) were calculated for each calf.

Faecal samples were collected from each calf by retrieval from the rectum on day 5, 7, 14, 21, 28, 35, 42, 49, and 56. Faecal samples were scored in respect to consistency by the same researcher on all collection days according to the following system: 1 = watery, diarrhoea; 2 = soft, unformed; 3 = soft, formed; 4 = hard, formed; and 5 = hard, dry pellets. On day 5 and 56, each faecal sample was diluted 10 folds with distilled water for pH measurement. The mixture of faecal sample and distilled water was homogenized and faecal pH was immediately measured using an electronic pH meter (PT-10, Sartorius AG, Goettingen, Germany) fitted with a glass electrode. Faecal samples could be collected from 8 of 10 and 9 of 10 calves in CG and EG, respectively, on day 56 by retrieval from rectum using sterile gloves. Faecal samples were placed in sterile sampling bags and immediately transported to the laboratory for bacterial enumeration. One g of each faecal sample was homogenized with 9 mL of saline peptone water in a stomacher laboratory blender (Stomacher 80 Biomaster, Seward Inc., Port Saint Lucie, FL, USA). Subsequently, serial 10-fold dilutions were made in saline peptone water and plated onto relevant selective media. *Lactobacillus* was grown on Man Rogosa Sharpe agar (MRS agar, Hypet Media, Diatek, Istanbul, Turkey). Each plate was incubated at 37°C for 48 h in an anaerobic

jar (Oxoid AN0035A, Basingstoke, Hampshire, UK) with gas generating sachet (Oxoid CN0020C, Basingstoke). Non-spore-former rods, gram-positive and catalase-negative isolates were regarded as *Lactobacillus*. *Bifidobacterium* was grown on Bifidobacterium Selective Medium agar (BSM agar, Hypet Media, Diatek). The plates were incubated at 37°C for 48 to 72 h in an anaerobic jar with gas generating sachet. The selected bacterial colonies were investigated with regard to cell morphology by Gram staining. The colonies with gram-positive rods and characteristic bifurcated "Y" and "V" shapes were recorded as *Bifidobacterium*. *C. perfringens* was grown on 4-methylumbelliferyl phosphate-supplemented (MUP, Merck 1.00888, Darmstadt, Germany) Tryptose Sulfite Cycloserine agar (TSC agar, Hypet Media, Diatek) containing egg yolk emulsion and selective supplement. Each plate was incubated at 37°C for 24 h in an anaerobic jar with gas generating sachet. Each presumptive black colony was added to 10 mL of Thioglycolate Broth (Merck 1.08190) and then incubated at 37°C for 16 to 18 h. Activated cultures were prepared for identifying by characteristic colony morphology, Gram staining and biochemical tests. The colonies with gram-positive and nonmotile rods, lactose positive, nitrate reduction positive, gelatine positive and motility negative were considered to be *C. perfringens*. Total coliform was grown on Violet Red Bile (VRB, Oxoid CM0107, Basingstoke) agar. After 24 to 48 h of incubation at 37°C, the red colonies with halos were typical for *E. coli*. The colonies typical for *E. coli* were enumerated and inoculated into Lactose Broth (LB, Oxoid CM0137, Basingstoke) with Durham tube. After 24 to 48 h of incubation at 44°C, gas and lactic acid production positive tubes were streaked onto Eosin Methylene Blue (EMB, Oxoid CM0069, Basingstoke) agar and confirmed to be *E. coli* by biochemical tests, such as Indol (SIM, Merck 1.05470), Methyl red and Voges-Proskauer (MR-VP, Merck 1.05712,) and Citrate (Oxoid CM0155, Basingstoke). Indol positive, MR positive, VP negative and citrate negative indicated the presence of *E. coli* type-1. The bacterial counts were expressed as log₁₀ CFU (colony forming unit) per gram of faecal samples.

Body temperatures were taken per rectum using a digital thermometer at the same times and recorded on day 5, 7, 14, 21, 28, 35, 42, 49, and 56. Health status of calves was monitored daily during the study. Faecal score of one was considered to be diarrhoea. In addition, tail and/or hind limbs stained with faeces were evaluated as a finding of diarrhoea. Calves with diarrhoea were treated by the farm veterinarian according to protocols established by the Department of Internal Medicine, Faculty of Veterinary Medicine. In the case of calves with diarrhoea, medication used (antibiotics, anti-inflammatory, immun stimulant, and

electrolyte products) and dosage and the duration of treatments applied for diarrhoea were recorded throughout the study. Costs associated with diarrhoea treatments were calculated based on current costs for each product and daily dosage for each medication for individual calves.

Statistical analysis

All statistical analyses were conducted by using SPSS ver. 5.0 (SPSS Inc., Chicago, IL, USA) (SPSS, 2004). Data for body weight, ADFI, ADG, and feed efficiency were tested to determine normal distribution by Kolmogorov-Smirnov test. Body weight, ADFI, ADG, and feed efficiency were tested for homogeneity of variance and analysed by independent sample T-test. Data for faecal score and faecal pH were tested to determine normal distribution by Kolmogorov-Smirnov test and F-test, respectively. Faecal score, faecal pH, faecal bacterial populations and body temperature were analysed by independent sample T-test. The incidence of diarrhoea was analysed by chi-square test and Fisher exact test. The statistical analyses for the duration of treatment applied for diarrhoea and the costs associated with diarrhoea treatments were performed by Mann-Whitney test. Differences between groups were considered significant at $p \leq 0.05$.

RESULTS AND DISCUSSION

Body weight, ADG, ADFI, and feed efficiency measurements are presented in Table 1. There were no significant differences ($p > 0.05$) in body weight, ADG, ADFI, and feed efficiency between groups during the period from day 5 to 56 after birth. However, body weight at the end of the study, ADG and ADFI values were 3.70%, 6.66%, and 10.97% higher for the calves supplemented with MOS than control calves, respectively. Similarly, MOS supplementation led to the numerical increases in final body weight, ADG and ADFI in other studies with calves

Table 1. Effect of MOS supplementation on growth performance of calves

Item	Group		p-value
	CG ^a (n = 10)	EG ^b (n = 10)	
Initial body weight (kg)	36.50 ± 0.83	37.31 ± 1.45	0.635
Final body weight (kg)	59.97 ± 2.43	62.19 ± 2.21	0.512
ADG (g/d)	450 ± 40	480 ± 30	0.604
ADFI (g/d)	597.58 ± 46.49	663.14 ± 37.33	0.290
Feed efficiency ^c (g gain/g feed)	0.75 ± 0.04	0.72 ± 0.03	0.565

MOS, mannanoligosaccharides; CG, control group; EG, experimental group; ADG, average daily weight gain; ADFI, average daily starter concentrate feed intake.

Values are presented as mean ± standard error of the mean.

^a Control group.

^b Group supplemented with MOS.

^c Feed efficiency: ADG/ADFI.

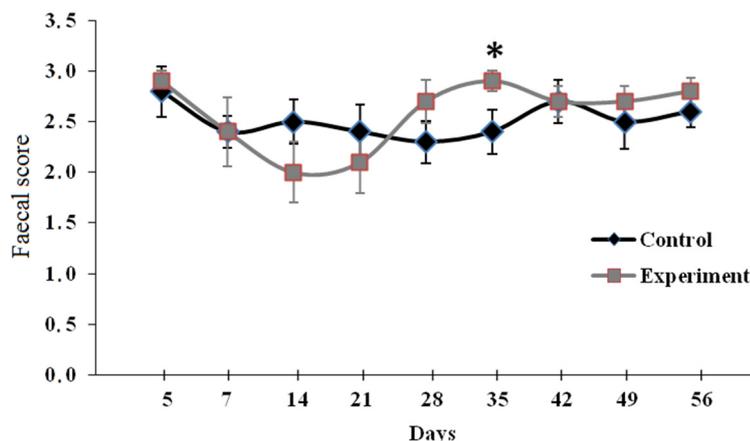


Figure 1. Mean (\pm standard error of the mean) faecal scores of control (CG, n = 10) and experimental (EG, n = 10) calves. CG, control group; EG, experimental group. * EG had a higher faecal score ($p = 0.05$) than CG on day 35.

(Uzmay et al., 2011; da Silva et al., 2012). Ghosh and Mehla (2012) observed that 4 g/d MOS supplementation improved ADG, ADFI, and feed efficiency of calves. White et al. (2002) reported that 3% MOS addition to diet decreased ADG and ADFI in weanling pigs. These different results of MOS on growth performance of livestock may be caused by level of supplementation of MOS from different sources and manufacturers, the duration of MOS supplementation, as well as nutrient composition of the diet, health status, intestinal bacterial populations and the environmental conditions (Demirel et al., 2007; Zhao et al., 2012).

In the current study, while faecal scores did not differ on day 5, 7, 14, 21, 28, 42, 49, and 56 between groups, EG had a higher faecal score ($p = 0.05$) than CG on day 35 (Figure 1). Based on the results for faecal score in our study, it can be suggested that MOS supplementation had no consistent effect on faecal score. Similarly, da Silva et al. (2012) also observed that 4 g/d MOS supplementation improved faecal score of calves only on week one during the experimental period lasting for eight weeks. Grieshop et al. (2004) reported that dietary supplementation with 1% MOS did not affect faecal score of dogs. Hill et al. (2008) observed no improvement in faecal score when 6 g/d of MOS was supplemented to calves. Morrison et al. (2010) reported that calves offered 10 g/d of MOS had an improved faecal score. In the study conducted by Ghosh and Mehla (2012), lower faecal score was observed in the calves supplemented with MOS (4 g/d) compared to the control calves. In contrast to our study, a higher faecal score would indicate formation of softer faeces in this study. Ghosh and Mehla (2012) suggested that the improved faecal score might be because of the suppression of pathogenic bacteria responsible for toxin production leading to intestinal secretion and diarrhoea (Giannella, 1983).

In the current study, faecal pH was not different ($p > 0.05$)

between groups (Table 2). Similarly, Swanson et al. (2002a) reported that supplementation of 2 g/d fructooligosaccharides plus 1 g/d MOS did not alter ileal and faecal pH of dogs. In the study conducted by Lebel et al. (2014), no effect of 0.1% MOS supplementation was observed on intestinal pH of pigs. Ao and Choct (2013) also demonstrated that caecal pH was not affected by dietary supplementation of MOS in broilers.

MOS is not enzymatically digested in the intestine and pathogenic bacteria bound to MOS are likely removed from the digestive system without attachment and colonization (Spring et al., 2000; Shane, 2001). Some studies in animals have shown that MOS supplementation can increase faecal *Lactobacillus* and/or *Bifidobacteria* concentrations (Swanson et al., 2002a; Grieshop et al., 2004) and decrease faecal *C. perfringens* or *E. coli* concentrations (Finuance et al., 1999; Grieshop et al., 2004). However, in other studies with MOS, no changes were observed in faecal bacterial populations (Swanson et al., 2002b; Terre et al., 2007). In our study, faecal concentration of *Lactobacillus* was lower ($p < 0.05$) in EG compared with CG and no differences ($p > 0.05$) in faecal concentrations of *Bifidobacterium*, *C. perfringens* and *E. coli* were found between CG and EG (Table 2). Grieshop et al. (2004) demonstrated that dietary supplementation with 1% MOS did not alter concentrations of faecal *Lactobacillus* and *C. perfringens* in dogs. However, Grieshop et al. (2004) reported that dogs supplemented with 1% MOS had a greater concentration of faecal *Bifidobacterium* compared with control dogs and MOS supplementation decreased concentration of faecal *E. coli*. Terre et al. (2007) observed that inclusion of 4 g/d MOS into the milk replacer did not affect faecal concentrations of *C. perfringens* and *E. coli* in calves. In the study conducted by Swanson et al. (2002a), supplementation of 2 g/d fructooligosaccharides plus 1 g/d MOS did not decrease faecal concentrations of *C.*

Table 2. Effect of MOS supplementation on faecal pH, faecal bacterial populations, incidence of diarrhoea, treatment days for diarrhoea and the costs associated with diarrhoea treatments in calves

Item	Group		p-value
	CG ^a (n = 10)	EG ^b (n = 10)	
Faecal pH			
Day 5	5.68±0.14	5.90±0.20	0.392
Day 56	7.37±0.12	7.46±0.08	0.514
Faecal bacterial populations ^c (CFU log ₁₀ /g fresh faeces)			
<i>Lactobacillus</i>	5.70±0.20	5.09±0.17	0.034
<i>Bifidobacterium</i>	5.28±0.40	5.59±0.14	0.461
<i>Clostridium perfringens</i>	4.15±0.41	3.74±0.20	0.336
<i>Escherichia coli</i>	4.27±0.56	4.70±0.36	0.508
The incidence of diarrhoea (%)	40.00	50.00	1.000
Treatment days/calf treated for diarrhoea	7.00±1.15	5.40±0.98	0.200
Cost of injectable antibiotic treatment (€/calf treated for diarrhoea)	2.77±0.69	1.76±0.58	0.190
Cost of total treatment (€/calf treated for diarrhoea) ^d	8.18±3.55	6.08±2.60	0.310

MOS, mannanoligosaccharides; CG, control group; EG, experimental group; CFU, colony forming unit.

Values are presented as mean±standard error of the mean unless otherwise indicated.

^a Control group. ^b Group supplemented with MOS.

^c Bacterial populations in sterile faeces sampled from 8 of 10 and 9 of 10 calves in CG and EG, respectively, on day 56.

^d Cost of injectable and oral antibiotics, anti-inflammatory, immun stimulant, and electrolyte treatments.

perfringens and *E. coli*. Swanson et al. (2002a) observed that faecal *Lactobacillus* and *Bifidobacterium* concentrations were greater in dogs supplemented with fructooligosaccharides plus MOS compared to control dogs. Based on the current study and aforementioned studies (Finuance et al., 1999; Swanson et al., 2002a,b; Grieshop et al., 2004; Terre et al., 2007), it can be reported that effects of MOS supplementation on intestinal bacterial populations are inconsistent. Lactate is a major end-product of the lactate-producing species, *Lactobacillus* and *Bifidobacterium*, and is an antimicrobial substance. Lactate-producing bacteria can decrease colonic pH by producing lactate, which can suppress the growth of pathogen populations in the intestine (Swanson et al., 2002a,b). In our study, decreased *Lactobacillus* concentration in EG may have led to a reduction in lactate production. No reduce in

concentrations of faecal *C. perfringens* and *E. coli* in EG may be partly associated with no decrease in faecal pH of the calves supplemented with MOS.

No difference ($p>0.05$) in body temperature was detected between groups throughout the study (Figure 2). Normal body temperature in calves ranges from 38.50°C to 39.50°C (Jackson and Cockcroft, 2002). Body temperature is a useful indicator of describing the health status of animals. Increased body temperature is an early sign of infection (Apanavicius et al., 2007). Calves with body temperature $>39.50^{\circ}\text{C}$ were considered to be febrile. There were four calves having a higher body temperature than 39.50°C in both CG and EG during the study.

Diarrhoea developed in 4 of 10 and 5 of 10 calves in CG and EG, respectively, during the study. No difference ($p>0.05$) in the incidence of diarrhoea was found between

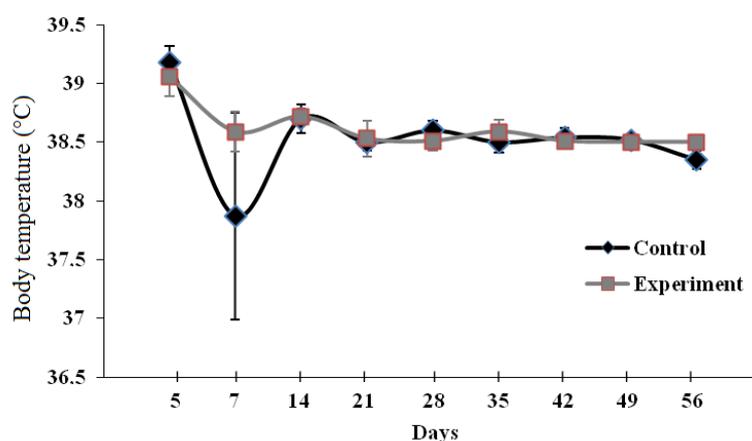


Figure 2. Mean (\pm standard error of the mean) body temperatures of control (CG, n = 10) and experimental (EG, n = 10) calves. CG, control group; EG, experimental group.

groups (Table 2). This result was supported by data from faecal score and faecal bacterial populations. A higher dose than 7 g/d of MOS supplement may be necessary to decrease faecal concentrations of pathogenic bacteria and the occurrence of diarrhoea under the circumstances where the incidence of diarrhoea is high as observed in our study. Throughout the study, respiratory and other health problems were not observed in any group. In this study, there were no significant differences ($p>0.05$) in the duration of treatments applied for diarrhoea (treatment days) and the costs associated with diarrhoea treatments between groups (Table 2). However, mean treatment days and costs of injectable antibiotic and total (injectable and oral antibiotics, anti-inflammatory, immun stimulant, and electrolyte products) treatments were numerically decreased in EG compared with CG. Magalhaes et al. (2008) also observed that treatment days with antibiotic and costs of total and antibiotic treatment were numerically decreased when calves were supplemented with yeast culture as a source of MOS. Uzman et al. (2011) reported that while cost of antibiotics used for treatment of diarrhoea was higher for calves supplemented with 4 g/d of MOS compared with control calves for day 5 to 25, antibiotic treatment cost in day 26 to 46 was decreased by MOS supplementation in the whole milk.

In conclusion, during the preweaning period, MOS supplementation in the whole milk did not significantly affect growth performance parameters in calves. Supplementation of MOS did not positively alter bacterial populations in faeces and did not significantly affect the incidence of diarrhoea, treatment days for diarrhoea and the costs associated with diarrhoea treatments. Higher doses than 7 g/d of MOS supplement may be necessary to decrease faecal concentrations of pathogenic bacteria and the occurrence of diarrhoea under the conditions of the present study where the incidence of diarrhoea is high. The observed decreases in treatment days and the cost of diarrhoea treatments for the calves supplemented with MOS were accompanied by increases in final body weight (3.70%), ADG (6.66%) and ADFI (10.97%), collectively, which may indicate potential benefit of MOS in treatment of diarrhoea in calves.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

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