



The Effects of Fat-soluble Vitamin Administration on Plasma Vitamin Status of Nursing Pigs Differ When Provided by Oral Administration or Injection

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ABSTRACT: Four experiments were conducted to investigate the effect of fat-soluble vitamin administration to sows or newborn pigs on plasma vitamin status. In Exp. 1 and 2, a total of 24 and 43 newborn pigs were allotted to control and vitamin treatments (vitamin D₃ with variable addition of vitamins A and E) orally or by i.m. injection. In Exp. 3, pigs from Exp. 2 were allotted to 2 treatments (\pm vitamins D₃ and E in drinking water) for 14 d postweaning. In Exp. 4, twenty-four gestating sows were used for 2 treatments (\pm injection of a vitamin D₃/A/E product 2 wk prepartum). In Exp. 1 and 2, when vitamin D₃ was administered orally or by i.m. injection on d 1 of age, pigs had increased plasma 25-hydroxycholecalciferol (25-OH D₃) concentration 10 d after administration compared with control pigs ($p < 0.05$). The injectable administration with vitamin D₃ and E was able to achieve higher plasma 25-OH D₃ ($p < 0.05$) and α -tocopherol ($p < 0.05$) concentrations than oral administration. At weaning, the pigs in the injection group had higher plasma 25-OH D₃ concentration than those in the other groups in both studies ($p < 0.05$). In Exp. 3, water supplementation of vitamin D₃ and E postweaning increased plasma 25-OH D₃ and α -tocopherol concentrations at d 14 postweaning ($p < 0.01$). In Exp. 4, when sows were injected with the vitamin D₃ product prepartum, serum 25-OH D₃ concentrations of sows at farrowing ($p < 0.01$), and in their progeny at birth ($p < 0.01$) and weaning ($p < 0.05$) were increased. These results demonstrated that fat-soluble vitamin administration to newborn pigs increased plasma 25-OH D₃ concentration regardless of administration routes and α -tocopherol concentration by the injectable route, and that water supplementation of vitamin D₃ and E to nursery pigs increased plasma 25-OH D₃ and α -tocopherol concentrations. Additionally, injecting sows with vitamin D₃ prepartum increased 25-OH D₃ in sows and their offspring. If continued research demonstrates that the serum levels of 25-OH D₃ are critical in weanling pigs, a variety of means to increase those levels are available to swine producers. (**Key Words:** Fat-soluble Vitamin, Piglets, Sows, Administration Routes, Vitamin D₃)

INTRODUCTION

Vitamin D has not been a major issue in the swine industry for several decades but in the past few years, attention on vitamin D and its status in pigs has risen with the increasing occurrence of metabolic bone diseases in some production settings (Madson et al., 2012). Recently, seemingly low concentrations of plasma vitamin D have been identified in periweaning failure to thrive syndrome

(PFTS), noted by veterinarians and researchers since 2008 (Huang et al., 2011). Consequently, products are available in the US as a means to specifically modify the plasma vitamin D levels in an attempt to prevent pigs from developing metabolic bone diseases and the mortality associated with PFTS. Some of the products contain only vitamin D while other products are a combination of fat-soluble vitamins.

Several recent studies have been conducted to investigate vitamin D requirements and/or supplementation/administration to sows and piglets (Lauridsen et al., 2010; Witschi et al., 2011; Rortvedt et al., 2012; Flohr et al., 2014a). The supplementation of vitamin D to gestation and lactation diets at concentrations above

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NRC (1998) requirement was demonstrated to increase serum vitamin D concentration of sows (Lauridsen et al., 2010) and their progeny (Witschi et al., 2011). Regarding vitamin D supplementation to piglets, when newborn pigs were supplemented with vitamin D₃ via oral administration at 40,000 or 80,000 IU of vitamin D₃, higher serum 25-hydroxycholecalciferol (25-OH D₃) concentration was detected at weaning compared with pigs without vitamin D₃ administration (Flohr et al., 2014a). On the other hand, Goff et al. (1984) suggested that the parenteral vitamin D₃ treatment to sows at d 20 prepartum by i.m. injection of vitamin D₃ was an effective method for enhancing vitamin D status of newborn piglets because the vitamin D status of sows was closely correlated to that of the fetus and neonatal piglets. Based on these limited results, an increased serum vitamin D concentration of piglets seems to be possible via several methods such as oral administration to piglets, supplementation to maternal feed and injection to sows. However, the relationship of that increased serum concentration to increased performance or bone measure is not always present. Regarding other fat soluble vitamins, vitamin E injection of 300 IU to neonatal pigs has been demonstrated to increase plasma α -tocopherol levels on 1 d after injection (Hill et al., 1999), and Håkansson et al. (2001) reported that plasma retinol concentration of suckling piglets were influenced by the vitamin A status of sows and colostrum. However, limited information is available for a direct comparison of fat-soluble vitamin administration to newborn pigs by different administration routes. Therefore, the objective of the current research was to evaluate the effects of a variety of supplemental fat-soluble vitamin products administered by different administration routes to young pigs as well as the efficacy of vitamin D₃ injection to sows prepartum.

MATERIALS AND METHODS

All procedures in this study were conducted in accordance with guidelines stated in the Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching by the Federation of Animal Science Societies (FASS, 2010).

Animals and experimental design

Exp. 1: A total of 24 pigs (Yorkshire×Duroc) were used from 4 litters (i.e., 6 pigs/sow; average parity: 4.0). At birth, pigs were assigned to 3 treatments within each litter based on gender and body weight (BW) in a randomized complete block design (RCBD). Treatments were: i) control: no supplemental vitamin, ii) i.m. injection of 0.8 mL of a vitamin complex (VITAL E-NEWBORN containing 500 IU of vitamin E as d- α -tocopherol, 50,000 IU of vitamin A as retinyl palmitate, and 50,000 IU of vitamin D₃ per mL), and

iii) oral administration of 0.8 mL of a vitamin complex (EMCELLE NEWBORN EAD containing 500 IU of vitamin E as d- α -tocopherol, 50,000 IU of vitamin A as retinyl palmitate, and 50,000 IU of vitamin D₃ per mL). The commercial products used were from Stuart Products Inc. (Bedford Texas, USA).

Exp. 2: This experiment was conducted to verify the effect of various combinations of oral fat-soluble vitamin product administration with control and injectable administration. A total of 43 pigs (Yorkshire×Duroc) were used from 4 litters (average parity: 4.8). At birth, a pig was assigned to each of 6 treatments within each litter in a completely randomized design (CRD); and, following the initial assignment of pigs, any remaining pigs in each litter were allotted to treatments 1 and 2. Treatments were: i) control: no supplemental vitamin, ii) i.m. injection of 1.00 mL of a vitamin complex (VITAL E-NEWBORN containing 500 IU of vitamin E as d- α -tocopherol, 50,000 IU of vitamin A as retinyl palmitate, and 50,000 IU of vitamin D₃ per mL), iii) oral administration of 1.00 mL of a vitamin complex (EMCELLE NEWBORN EAD containing 500 IU of vitamin E as d- α -tocopherol, 50,000 IU of vitamin A as retinyl palmitate, and 50,000 IU of vitamin D₃ per mL), iv) oral administration of 1.66 mL of a vitamin D and E complex (EMCELLE ED3 containing 30,000 IU of vitamin D₃ and 500 IU of vitamin E as d- α -tocopherol per mL), v) oral administration of 0.60 mL of vitamin D₃ (EMCELLE D3 containing 84,500 IU of vitamin D₃ per mL), and vi) oral administration of 1.66 mL of vitamin E (EMCELLE E containing 500 IU of vitamin E as d- α -tocopherol per mL). The products used were from Stuart Products Inc. (Bedford Texas, USA).

Exp. 3: All piglets from Exp. 2 were allotted to 1 of 2 additional treatments at weaning in a split plot design. With a main plot from the treatments at birth in Exp. 2, a subplot was divided into a control group without a vitamin product in drinking water and a group that consumed drinking water with a vitamin D₃ and E product (EMCELLE ED3-LIQUID, Stuart Products Inc., Bedford, Texas, USA) containing 30,000 IU of vitamin D₃ and 500 IU of vitamin E as d- α -tocopherol per mL during the first 14 d after weaning. The product was proportioned into the water at a rate that provided drinking water that contained 5,658 IU of vitamin D₃ and 95 IU of vitamin E per L.

Exp. 4: A total of 24 sows (Pig Improvement Company; average parity: 5.2) were allotted to 2 treatments in a CRD. Treatments were divided into a control group without injection and a group that received a 5 mL injection at 2 wk prepartum of a vitamin D₃ product (VITAL E-Hi A+D, Stuart Products Inc., Bedford, Texas, USA) containing 100,000 IU of vitamin D₃, 300 IU of vitamin E as d- α -tocopherol, and 200,000 IU of vitamin A as retinyl

palmitate per mL.

Diets, housing and sample collection

Exp. 1 and 2: All sows with pigs were kept in individual farrowing crates in an environmentally controlled farrowing facility without windows. In Exp. 1, the lactation diet for sows was a corn-soybean meal (SBM)-based diet. In Exp. 2, the lactation diets for sows were corn-SBM-based diets with 10% distillers dried grains with solubles (DDGS). In both experiments, the gestation and lactation diets contained the following per kg diet: vitamin A, 6,600 IU; vitamin D₃, 880 IU; vitamin E, 44 IU; vitamin K (menadione sodium bisulfite complex), 6.6 mg; riboflavin, 8.8 mg; d-pantothenic acid, 22 mg; niacin, 44 mg; vitamin B₆, 4.4 mg; vitamin B₁₂, 33 µg; d-biotin, 220 µg; and folic acid, 1.32 mg. Sows were provided 1.8 to 2.5 kg of the gestation diet before being brought to the farrowing rooms at about d 112 of gestation. Sows were provided the lactation diet *ad libitum* and water was freely available from a water nipple throughout the experimental period. All pigs were processed at birth (within 15 h) and assigned to a treatment. Processing of the piglets involved weighing, ear-notching, needle teeth clipping, and iron injection with 100 mg as Fe-dextran. With regard to administration of treatments, pigs were administered vitamin products by either injection or oral placement in the back of the mouth. Injectable products were provided to each pig in the trapezius muscle on the opposite side of the neck where the iron injection was given. Orally administered treatments were provided through a plastic tube attached to a 3 mL syringe into which the proper dosage had been drawn. The BW of the pigs were also recorded about 10 d after administration (average 11.75 d of age, 12±1 d for Exp. 1; average 12.00 d of age, 12±1 d for Exp. 2) and weaning (19.50 d of age, 20±1 d for Exp. 1; 20.25 d of age, 20±1 d for Exp. 2) to calculate growth performance. In addition, the BW of sows was obtained at farrowing and weaning.

Blood samples of sows were taken into heparin-containing tubes from the anterior vena cava at farrowing and weaning. Blood samples of pigs were collected from the anterior vena cava on d 0 or 1 (before administration of any treatments), approximately 10 d after administration of treatments and weaning. Blood samples were centrifuged at 1,700 g for 15 min at 4°C; plasma samples were then aliquoted into microtubes and stored at -20°C until analysis. Plasma samples were sent to the Iowa State University Veterinary Diagnostic Laboratory for vitamin assay (Ames IA, USA).

Exp. 3: Five to seven pigs were housed per 1.22×1.22 m² raised-deck nursery pen with plastic coated expanded metal flooring. The diet provided was a 1.21% standardized ileal digestible lysine corn-SBM-based starter diet that

contained the following per kg diet: vitamin A, 4,950 IU; vitamin D₃, 660 IU; vitamin E, 33 IU; vitamin K (menadione sodium bisulfite complex), 4.95 mg; riboflavin, 6.6 mg; d-pantothenic acid, 16.5 mg; niacin, 33 mg; vitamin B₆, 3.3 mg; vitamin B₁₂, 24.75 µg; d-biotin, 165 µg; and folic acid, 990 µg.

Blood samples of pigs were collected from the anterior vena cava at d 14 postweaning. Blood samples were centrifuged at 1,700 g for 15 min at 4°C, aliquoted into microtubes and stored at -20°C until analysis. Plasma samples were sent to the Iowa State University Veterinary Diagnostic Laboratory for vitamin assay. Blood data at weaning from Exp. 2 were used as the initial values in Exp. 3.

Exp. 4: All sows with pigs were housed in individual farrowing crates in an environmentally controlled farrowing facility. The sows were provided a common gestation diet containing 2,750 IU/kg of vitamin D₃, as well as a common lactation diet that also contained 2,750 IU/kg of vitamin D₃. The gestation diet was a corn-SBM-based diet with 60% DDGS and phytase (750 FTU/kg; Phyzyme XP 10000, Danisco Animal Nutrition, Marlborough, UK) and the lactation diet was a corn-SBM-based diet with 20% DDGS and phytase (750 FTU/kg; Phyzyme XP 10000, Danisco Animal Nutrition). Water was freely available throughout the experimental period.

The injectable product was administered into a neck muscle and sows were bled immediately preceding administration of the product and again at parturition. Two piglets were selected from each litter at birth for blood sampling; one of the two selected piglets was bled again at weaning. Blood samples were centrifuged; serum samples were put into microtubes and submitted to the Iowa State University Veterinary Diagnostic Laboratory for analysis of 25-OH D₃ concentrations.

Statistical analysis

Data from Exp. 1 were analyzed by ANOVA for a RCBD using MIXED procedure of SAS (SAS Inst. Inc., Cary, NC, USA). Fixed effect was the vitamin treatment and random effects were the litter, gender and gender× treatment interaction. Data from Exp. 2 were analyzed by ANOVA for a CRD using MIXED procedure of SAS. Fixed effect was solely the vitamin treatment. Data of plasma vitamin levels of sows between farrowing and weaning were analyzed by ANOVA using MIXED procedure of SAS containing the day (farrowing and weaning) as a fixed effect and the sow as a random effect. In Exp. 3, pigs from treatments 1 and 2 from Exp. 2 were used for statistical analysis whereas treatments 3 to 6 from Exp. 2 were not used in the analysis because of inadequate replication and imbalance among treatments. Data from Exp. 3 were analyzed by ANOVA for a split plot design using MIXED

procedure of SAS. Main plot was lactation treatment and subplot was nursery water treatment. Models included lactation treatment, nursery water treatment, and their interaction as fixed effects, and the pig within lactation treatment as a random effect. Data from Exp. 4 were analyzed by ANOVA for a CRD using MIXED procedure of SAS. Fixed effect was solely the vitamin treatment. Individual pig and sow were considered the experimental units. Least squares means were separated using PDIF option in SAS. Statistical differences were considered significant at $p < 0.05$, and a trend at $p < 0.10$.

RESULTS

The effect of vitamin D₃ supplementation with vitamins A and E to newborn pigs in Exp. 1 on plasma 25-OH D₃, retinol, and α -tocopherol concentrations is shown in Table 1. At 10 d after administration all groups treated with a vitamin D₃ product had higher plasma 25-OH D₃ concentration than the control group ($p < 0.05$). Additionally, when the pigs received the injectable product, the plasma 25-OH D₃ concentration was the highest among treatments ($p < 0.05$). At weaning, plasma values for injection and oral treatments declined from 10 d after administration but plasma values for the injection treatment remained elevated relative to plasma values for the other pigs ($p < 0.05$). With regard to plasma retinol and α -tocopherol concentrations, there were no differences except at 10 d after administration when plasma α -tocopherol concentrations of pigs in the injection group were the highest among all treatments

($p < 0.05$) and tended to be maintained until weaning ($p = 0.09$).

In Exp. 2 (Table 2), the effect of vitamin D₃ and variable vitamin A and E administration to newborn pigs on plasma 25-OH D₃ concentrations demonstrates again that the plasma concentrations at 10 d after administration can be increased by either oral or injectable administration ($p < 0.05$), that values decline from 10 d after administration to weaning regardless of treatment, and that values at weaning can remain higher than those of control pigs in which no treatment was administered ($p < 0.05$). The injectable product resulted in the highest plasma values and the results were higher than the oral administration treatments at weaning ($p < 0.05$). In the case of α -tocopherol, the vitamin E injection to piglets was also more effective to increase plasma α -tocopherol concentration at 10 d after administration compared with the oral administration ($p < 0.05$). However, at weaning the highest numerical value was actually from one of the oral products that contained vitamin E and it, as well as the injectable product, was higher than that of the control pigs ($p < 0.05$). Plasma 25-OH D₃ levels of sows were higher at weaning than farrowing in both Exp. 1 and 2 ($p < 0.05$).

The effects of the treatments on BW and growth in Exp. 1 and 2 are provided in Table 3 and 4. No differences among treatments were detected on BW and average daily gain (ADG) of pigs during suckling periods in either experiment.

The effect of vitamins D₃ and E supplementation via drinking water to nursery pigs in Exp. 3 is presented in

Table 1. Effect of fat-soluble vitamin administration to pigs on plasma 25-hydroxycholecalciferol (25-OH D₃), retinol, and α -tocopherol concentration (Exp. 1)^{1,2}

Criteria	Sow (n = 4)	Treatments ³			SEM	p-values
		Control	Inj. vit. A, D ₃ , E	Oral vit. A, D ₃ , E		
25-OH D ₃ (ng/mL)						
Initial	21.23 ⁴	2.58	2.68	2.65	0.18	0.92
d 10		5.87 ^c	82.61 ^a	38.49 ^b	3.33	<0.01
Weaning	31.60	5.88 ^b	30.53 ^a	10.01 ^b	3.26	0.04
Retinol (μ g/mL)						
Initial		0.16	0.13	0.14	0.02	0.50
d 10		0.37	0.37	0.28	0.05	0.32
Weaning		0.35	0.20	0.19	0.08	0.16
α -Tocopherol (μ g/mL)						
Initial		2.25	1.83	1.83	0.35	0.62
d 10		6.85 ^b	15.20 ^a	3.75 ^b	1.91	0.01
Weaning		3.93	5.93	3.62	0.93	0.09

SEM, standard error of the mean.

¹ 24 pigs were used for 25-OH D₃ analysis (n = 8 for each treatment) and 12 pigs were used for retinol and α -tocopherol analysis (n = 4 for each treatment).

² Initial, d 0 or 1 of age before administration of treatments; d 10, 10 d after administration (average 11.75 d of age); weaning, average 19.50 d of age.

³ The injectable and oral products supplied 400 IU vitamin E, 40,000 IU vitamin A, and 40,000 IU vitamin D₃ and were administered on d 1 of life.

⁴ There was a difference in 25-OH D₃ levels of sows between farrowing and weaning ($p < 0.05$; SEM: 2.10).

^{a,b,c} Means in the same row without a common superscript differ ($p < 0.05$).

Table 2. Effect of fat-soluble vitamin administration to pigs on plasma 25-hydroxycholecalciferol (25-OH D₃), retinol, and α -tocopherol concentration (Exp. 2)^{1,2}

Criteria	Sow (n = 4)	Treatment ³						SEM	p-values
		Control	Inj. A, D ₃ , E	Oral A, D ₃ , E	Oral D ₃ , E	Oral D ₃	Oral E		
25-OH D ₃ (ng/mL)									
Initial	26.45 ⁴	4.41	4.47	4.40	5.45	4.38	5.10	0.82	0.93
d 10		8.25 ^b	110.74 ^a	77.17 ^a	84.68 ^a	86.20 ^a	6.47 ^b	15.28	<0.001
Weaning	32.65	5.02 ^c	40.24 ^a	26.10 ^b	27.30 ^b	30.88 ^b	5.10 ^c	3.13	<0.001
Retinol (μ g/mL)									
Initial	0.16	0.04	0.07	0.11	0.08	0.10	0.10	0.02	0.21
d 10		0.23	0.25	0.25	0.21	0.26	0.28	0.03	0.69
Weaning	0.27	0.18	0.16	0.17	0.16	0.11	0.18	0.05	0.92
α -Tocopherol (μ g/mL)									
Initial	1.23 ⁴	3.24	2.86	3.68	4.30	2.70	1.43	1.38	0.88
d 10		4.78 ^b	11.26 ^a	5.37 ^b	6.93 ^{ab}	4.80 ^b	5.80 ^b	1.59	<0.01
Weaning	3.13	4.51 ^b	7.16 ^a	5.93 ^{ab}	7.33 ^a	3.85 ^b	4.80 ^{ab}	0.84	<0.01

SEM, standard error of the mean.

¹ 43 pigs were used for all analysis (n = 14, 13, 4, 4, 4, and 4 for control, Inj. A, D₃, E; Oral A, D₃, E; Oral D₃, E; Oral D₃; Oral E).² Initial, d 0 or 1 of age before administration of treatments; d 10, 10 d after administration (average 12.00 d of age); weaning, average 20.25 d of age.³ See text for more complete description of treatments.⁴ There were differences in 25-OH D₃ and α -tocopherol levels of sows between farrowing and weaning (p<0.05; SEM: 2.96, 0.07, and 0.16 for 25-OH D₃, retinol and α -tocopherol, respectively).^{a,b,c} Means in the same row without a common superscript differ (p<0.05).**Table 3.** Effect of fat-soluble vitamin administration to pigs on growth performance (Exp. 1)^{1,2}

Criteria	Sow (n = 4)	Treatment ³			SEM	p-values
		Control	Inj. vit. A, D ₃ , E	Oral vit. A, D ₃ , E		
BW (kg)						
Initial	231.03	1.73	1.58	1.53	0.18	0.43
d 10		4.21	3.99	4.06	0.37	0.87
Weaning	242.02	6.34	5.98	6.20	0.53	0.86
ADG (kg/d)						
Initial to d 10		0.21	0.20	0.21	0.02	0.92
d 10 to weaning		0.27	0.26	0.28	0.02	0.76
Initial to weaning		0.24	0.22	0.24	0.02	0.88

SEM, standard error of the mean; BW, body weight; ADG, average daily gain.

¹ 24 pigs were used (n = 8 for each treatment).² Initial, d 0 or 1 of age before administration of treatments; d 10, 10 d after administration (average 11.75 d of age); weaning, average 19.50 d of age.³ The injectable and oral products supplied 400 IU vitamin E, 40,000 IU vitamin A, and 40,000 IU vitamin D₃ and were administered on d 1 of life.**Table 4.** Effect of fat-soluble vitamin administration to pigs on growth performance (Exp. 2)^{1,2}

Criteria	Sow	Treatment ³						SEM	p-values
		Control	Inj. A, D ₃ , E	Oral A, D ₃ , E	Oral D ₃ , E	Oral D ₃	Oral E		
BW (kg)									
Initial	229.5	1.56	1.55	1.51	1.60	1.58	1.83	0.17	0.86
d 10		3.51	3.27	3.84	3.05	3.87	4.37	0.45	0.47
Weaning	248.2	5.53	5.12	5.84	4.12	6.44	6.90	0.72	0.24
ADG (kg/d)									
Initial to d 10		0.17	0.16	0.19	0.14	0.21	0.24	0.03	0.31
d 10 to weaning		0.25	0.23	0.25	0.14	0.31	0.28	0.04	0.22
Initial to weaning		0.20	0.19	0.21	0.14	0.25	0.26	0.03	0.25

SEM, standard error of the mean; BW, body weight; ADG, average daily gain.

¹ 43 pigs were used for all analysis (n = 14, 13, 4, 4, 4, and 4 for control, Inj. A, D₃, E; Oral A, D₃, E; Oral D₃, E; Oral D₃; Oral E).² Initial, d 0 or 1 of age before administration of treatments; d 10, 10 d after administration (average 12.00 d of age); weaning, average 20.25 d of age.³ See text for more complete description of treatments.

Table 5. Effect of fat-soluble vitamin administration by injection in early lactation and drinking water to pigs on plasma 25-hydroxycholecalciferol (25-OH D₃), retinol, and α -tocopherol concentration (Exp. 3)¹

Treatment at birth	Control		Injection A, D ₃ , E		SEM	p-values		
	Without	With	Without	With		Con vs Inj.	Water	Interaction
No. of pigs	6	7	6	5				
25-OH D ₃ (ng/mL)								
At weaning	6.07	4.11	43.55	37.17	2.96	<0.001	0.10	0.36
d 14 postweaning	33.57	65.91	38.95	68.08	2.67	0.27	<0.001	0.43
Change ²	27.07	61.80	-4.60	31.10	3.39	<0.001	<0.001	0.87
Retinol (μ g/mL)								
At weaning	0.24	0.13	0.18	0.15	0.04	0.68	0.15	0.37
d 14 postweaning	0.31	0.22	0.29	0.22	0.03	0.71	0.01	0.75
Change ²	0.07	0.09	0.11	0.06	0.06	0.94	0.77	0.55
α -Tocopherol (μ g/mL)								
At weaning	4.28	4.78	6.28	8.22	0.67	<0.01	0.11	0.31
d 14 postweaning	0.87	2.87	1.72	3.05	0.24	0.07	<0.001	0.17
Change ²	-3.42	-1.92	-4.57	-5.18	0.72	0.01	0.56	0.18

SEM, standard error of the mean.

¹ Treatment: Control, Treatment 1 from Exp. 2; Inj. A, D₃, E, Treatment 2 from Exp. 2; With, drinking water containing vitamin D₃ (5,658 IU/L) and E (95 IU/L) during 14 d after weaning; Without, no vitamin supplementation in drinking water.² Change was calculated by subtracting values at weaning from values at d 14 postweaning.

Table 5. The control and injection treatments from the nursing phase in Exp. 2 were used as the main treatment effects in the statistical analysis of Table 5. When pigs then received vitamins D₃ and E via drinking water in the nursery, the pigs had higher plasma 25-OH D₃ and α -tocopherol concentrations at d 14 postweaning ($p < 0.01$). Additionally, the change of plasma 25-OH D₃ concentration from weaning to d 14 postweaning in pigs supplemented with vitamins via drinking water was also higher than pigs without water vitamin supplementation ($p < 0.01$). In contrast, plasma retinol concentration was adversely affected at d 14 postweaning by high levels of vitamins D₃ and E in drinking water ($p < 0.05$). Even though there was no difference on plasma 25-OH D₃ concentration at d 14 postweaning between the lactation treatments (control and injection), a lower change of plasma value was observed in the injection treatment that had higher plasma values at weaning compared with the control treatment ($p < 0.01$). In the case of plasma α -tocopherol concentration, higher plasma values were observed in the injection treatment than the control treatment at weaning ($p < 0.01$) and a trend at d 14 postweaning ($p = 0.07$). The plasma values declined from weaning to d 14 postweaning and a greater reduction was detected in the injection treatment ($p < 0.05$).

In Exp. 4 (Table 6), a vitamin D₃/A/E injection to gestating sows increased absolute serum 25-OH D₃ concentration at farrowing and its change from pre-administration values ($p < 0.01$). Piglets from sows that received the prepartum vitamin D₃ injection had higher serum 25-OH D₃ concentrations than those in the control group at birth ($p < 0.01$) and this improvement was

maintained until weaning ($p < 0.05$) but vitamin D₃ injection to sows did not affect the change of serum 25-OH D₃ concentration of piglets during suckling period.

DISCUSSION

This is the first study to investigate the plasma fat-soluble vitamin profiles of nursing piglets administered with fat-soluble vitamins by different administration routes.

Table 6. Effect of vitamin D₃ injection to gestating sows on serum 25-hydroxycholecalciferol concentration of sows and piglets (Exp. 4)^{1,2}

Criteria	Control	Vitamin D ₃ injection	SEM	p-values
Sows (ng/mL)				
Initial ³	42.07	37.10	2.61	0.19
At farrowing	43.10	58.60	2.86	<0.01
Change ⁴	1.03	21.50	3.19	<0.001
Piglets (ng/mL)				
At birth	4.00	6.15	0.36	<0.001
At weaning	9.67	12.05	0.81	0.05
Change ⁵	5.67	5.53	0.80	0.90

SEM, standard error of the mean.

¹ 24 sows were used (n = 11 and 13 for control and injection treatments, respectively).² 48 piglets at birth (n = 22 and 26 for control and injection treatments, respectively); 23 piglets at weaning and for change (n = 11 and 12 for control and injection treatments, respectively).³ Before injection at 2 wk prepartum.⁴ Change was calculated by subtracting values at initial from values at farrowing.⁵ Change was calculated by subtracting values at birth from values at weaning.

In both Exp. 1 and 2, the results, collectively, demonstrate that the vitamin D₃ status of pigs at weaning can be improved by administration of vitamin D₃ by a variety of methods and the results agree with those of recent studies (Rortvedt et al., 2012; Flohr et al., 2014a). However, plasma 25-OH D₃ and α -tocopherol concentrations declined from 10 d after administration to weaning in pigs from the injection and oral treatments, which agrees with Flohr et al. (2014a). This result suggests that plasma values are presumably diluted with the increase in blood volume associated with the rapid growth of the pigs as well as utilized by, or deposited in, various tissues. In contrast to Exp. 2, there was no decline in plasma 25-OH D₃ level of control pigs in Exp. 1. Because of no vitamin administration to the control pigs, the control pigs illustrate the normal physiological response in the suckling period but the reason for the discrepancy between experiments is unclear. Therefore, further study is needed to verify these responses.

Plasma 25-OH D₃ levels in sows increased from farrowing to weaning in both studies which may result from the feed intake change from the restricted feeding in gestation to *ad libitum* in lactation. Flohr et al. (2014b) reported sows fed gestation and lactation diets containing vitamin D₃ had increased 25-OH D₃ levels in plasma and milk by increasing supplementation levels from 1,500 to 6,000 IU per kg diet at farrowing, 10 d of lactation and weaning but there were no day effect and interaction between treatment and day of lactation in milk 25-OH D₃ levels. Therefore, this result indicates that, while plasma 25-OH D₃ levels in sows can be increased from farrowing to weaning by absolute higher vitamin D₃ intake in lactation than gestation, milk 25-OH D₃ levels for the suckling piglet may be more difficult to increase. However, it should be noted from the current study that plasma values for control pigs were higher at 10 d after administration than at birth which demonstrates that milk clearly contributes to the vitamin D status of the pig.

Comparing the results between oral and injectable methods in the present experiments, it was demonstrated that the i.m. injection of vitamin D₃, and E was able to enhance those statuses more than the oral administration. There have been few studies to compare the efficacy of different vitamin administration routes to pigs. In the case of sheep, ewes that received vitamin D₃ by i.m. injection had higher plasma 25-OH D₃ concentration and more lasting effects than those treated by oral administration (Hidiroglou et al., 1984). The parenteral administration of vitamin D₃ does not require the process of intestinal absorption that oral administration of vitamin D₃ requires. It has been documented that only 50% of orally administered vitamin D is absorbed (McDowell, 2000). Therefore, the process of intestinal absorption of vitamins probably

affected the difference of plasma 25-OH D₃ and α -tocopherol concentration in this study. Regarding vitamin A status, there was no effect on plasma retinol concentration of piglets by vitamin A administration. When dogs were administered 10,000 IU of retinyl palmitate per kg BW orally, plasma retinol concentration increased slightly to be maximized at 6 h, and then declined to the baseline at 48 h (Raila et al., 2002). These results suggest that each fat-soluble vitamin may have different biological responses and effectiveness to enhance plasma vitamin profiles by types and administration routes.

There were no differences among treatments on BW and ADG of pigs during the suckling period in either experiment. These results again agree with some recent research (Flohr et al., 2014a). Rortvedt et al. (2012) also reported no improvement of ADG for 35 d after weaning in pigs administered a single oral mega-dose of vitamin D₃ (40,000 IU) at birth (0.38 kg/d) compared with control pigs (0.41 kg/d). In contrast, Tousignant et al. (2013) reported that in field conditions, pigs administered vitamin D₃ (40,000 IU) orally at 2 d of age were heavier at weaning and d 7 postweaning than control pigs without vitamin D₃ administration but that the difference vanished at d 26 postweaning. The results of the current study indicated that there was no improvement in piglet growth by vitamin administration even though plasma levels of vitamins were clearly increased. This would seem to indicate that although some individuals may categorize normal plasma levels as "low", the levels are adequate for normal growth in an environment such as existed for the current research.

In Exp. 3, the water supplementation of vitamin D₃ and E increased plasma 25-OH D₃ and α -tocopherol concentrations which mean supplementation of vitamins via drinking water to nursery pigs is an effective method to enhance plasma status of vitamins. This result agreed with a previous study which reported the water supplementation of vitamin D₃ to nursery pigs from weaning for 10 d elevated serum 25-OH D₃ concentration on d 10 postweaning (Flohr et al., 2014a). For vitamin E status, Wilburn et al. (2008) reported the water vitamin E supplementation increased plasma α -tocopherol concentration of weanling pigs.

With regard to the changes of plasma 25-OH D₃ concentrations, the pigs in the lactation injection group that had higher plasma values at weaning had smaller increases of plasma 25-OH D₃ concentrations at d 14 in response to the water supplementation compared with those in the control group that had lower values at weaning, resulting in no difference in absolute plasma values between the lactation treatments at d 14 postweaning. Trang et al. (1998) reported there is an inverse linear correlation between basal 25-OH D₃ and the increase in 25-OH D₃ for vitamin D₃-treated humans. This result means that improvements from

vitamin treatment at birth could vanish after weaning when pigs are switched from milk to feed. In the case of plasma α -tocopherol concentration, the values declined after weaning regardless of treatment which is in line with the results from previous studies (Bonnette et al., 1990; Moreira and Mahan, 2002; Wilburn et al., 2008). On the other hand, even though a greater reduction of plasma values was observed in pigs from the injection group that had higher plasma values at weaning than those in the control group that had lower values, the pigs in the injection group still maintained higher plasma α -tocopherol levels than those in the control group. In previous studies, plasma α -tocopherol concentration declined steadily over 21 d after weaning when pigs were not supplemented with vitamin E (Moreira and Mahan, 2002; Wilburn et al., 2008). Plasma α -tocopherol concentration in the current study was measured only at d 14 postweaning which does not allow determination if that decline would continue; thus, the effect of injection at birth seems to remain. However, because of limited data available in this study and lack of previous research, further study is needed to explain the changes in plasma α -tocopherol concentration from weaning when pigs have different plasma levels at weaning.

In Exp. 4, a vitamin D₃ injection to gestating sows increased serum 25-OH D₃ levels of sows at farrowing and of their offspring at birth. This result means the maternal vitamin D status affects the status of offspring at birth and it may result from the placental transportation which agrees with the observations of Goff et al. (1984) who reported that when sows received vitamin D₃ by i.m. injection at d 20 prepartum, vitamin D status of their progeny was improved. Even though the difference in the serum 25-OH D₃ levels of piglets between treatments was maintained until weaning there was no difference in the magnitude of that treatment difference (or the change from birth to weaning) of 25-OH D₃ concentration in serum of piglets which means the improvement of vitamin D₃ status of gestating sows probably did not influence sow milk content. It may be noted that dietary vitamin D level was approximately 3 times higher in Exp. 4 than in Exp. 1 and 2, that the serum values of control sows at farrowing in Exp. 4 were approximately 2 times higher than those in Exp. 1 and 2, and that weaning serum values of control piglets in Exp. 4 were double that of the values at birth and 55% to 93% higher than the weaning values for control pigs in Exp. 1 and 2. Therefore, while milk may not be one of the best means of supplying vitamin D₃, it either is adequate to increase serum values of piglets if sows are supplemented at levels above NRC (1998) recommendation or else the colostrum D₃ concentration is elevated sufficiently from the sow dietary concentrations to allow the piglet serum values at weaning to be maintained at a higher level.

IMPLICATIONS

Vitamin D₃ supplementation to newborn pigs can improve vitamin D status by increasing plasma 25-OH D₃ concentrations regardless of administration routes. Vitamin A status of pigs was not affected by any of the means of administration whereas vitamin E status was affected by both routes of administration but more so by injection. Water supplementation of vitamin D₃ and E to nursery pigs can enhance vitamin D and E status by increasing plasma 25-OH D₃ and α -tocopherol concentrations and initial vitamin status of pig plasma at weaning may not affect its plasma status at 14 d after weaning. The improvement of vitamin D status of pregnant sows can result in the enhancement of its status of their offspring. Several products and modes of administration exist to modify vitamin D status of piglets if that is a matter of concern for a swine operation. Further studies are needed, however, to determine if this altered vitamin D status has value in different production settings.

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